Assessment effectiveness and safety of Thrombolytic Therapy to Pulmonary Emboli (PE) by Endobronchial Ultrasound-guided Transbronchial Needle Injection (EBUS-TBNI) in a Porcine Survival Model

This study has revealed that t-PA EBUS-TBNI can be an effective procedure to safely dissolve acute massive PE.
Assessment of Effectiveness and Safety of Thrombolytic Therapy to Pulmonary Emboli by Endobronchial Ultrasound-guided Transbronchial Needle Injection

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Glossary of Abbreviations:

PE: pulmonary embolism

USAT: ultrasound-accelerated thrombolysis therapy
Central Picture Legend (87/90 characters, including spaces)
Injection of t-PA using Endobronchial ultrasound-guided transbronchial needle injection

Central Message (182/200 characters, including spaces)
Injection of t-PA using the endobronchial ultrasound-guided transbronchial needle injection for pulmonary embolism can safely dissolve clots better than t-PA systemic administration.

Perspective Statement (328/405 characters, including spaces)
Acute pulmonary embolism at high risk remains a highly fatal disease. No major advances or changes have been made in the last decade. Our proposed method, using the endobronchial ultrasound-guided transbronchial needle injection, may be a major step toward improvement of treatment for the acute pulmonary embolism at high risk.
Structured Abstract (240/250 words)

Objective: Endobronchial ultrasound-guided transbronchial needle injection (EBUS-TBNI) may effectively treat acute pulmonary embolisms (PEs). Here, we assessed the effectiveness of clot dissolution and safety of t-PA injection using EBUS-TBNI in a one week survival study of a porcine PE model.

Methods: Six pigs with bilateral PEs were used: three for t-PA injection using EBUS-TBNI (TBNI group); three for systemic administration of t-PA (Systemic group). Once bilateral PEs were created, each 25 mg of t-PA injection using EBUS-TBNI for bilateral PEs (a total of 50 mg t-PA) and 100 mg of t-PA systemic administration were performed on day 1. Hemodynamic parameters, blood tests, and contrast-enhanced CT scans were carried out at several time points. On day 7, pigs were sacrificed to evaluate the residual clot volume in the pulmonary arteries.

Results: The average of percent change of residual clot volumes was significantly lower in TBNI group than in Systemic group (%: Systemic group 36.6 ± 22.6 vs TBNI group 9.6 ± 6.1, p < 0.01) on day 3. Considering the elapsed time, the average decrease of clot volume per hour at pre t-PA to post t-PA was significantly higher in TBNI group than in Systemic group (mm$^3$/hour: Systemic 68.1 ± 68.1 vs TBNI 256.8 ± 148.1, p < 0.05). No hemorrhage was observed intracranially, intrathoracically, or intraperitoneally on any contrast-enhanced CT images.

Conclusions: This study has revealed that t-PA injection using EBUS-TBNI is an effective and safe way to dissolve clots.

Keywords: endobronchial ultrasound, transbronchial needle injection, preclinical animal model, pulmonary embolism, thrombolysis
(Introduction)

Acute PE at high risk is associated with a high mortality rate; for patients with systolic blood pressure lower than 90 mmHg and/or hemodynamic instability lasting more than 15 minutes, the risk of 30-day mortality is about 20%. Currently, systemic thrombolytic therapy, rather than catheter-directed thrombolysis, is the suggested treatment for patients with acute PE at high risk.\textsuperscript{1}

\textsuperscript{3} The ultrasound-accelerated thrombolysis therapy (USAT) was reported as an improved therapy for PE\textsuperscript{4} and even though some of meta-analysis reported its usefulness\textsuperscript{5-7}, the evidence was insufficient to change the guidelines.\textsuperscript{3} There is room to increase the effectiveness of treatment of tissue plasminogen activator (t-PA) therapy as a treatment strategy for acute PE. In this study, we have examined the safety and the feasibility of endobronchial ultrasound (EBUS)-guided transbronchial needle injection (TBNI) as a means of improved t-PA delivery. EBUS is a well-established modality for the diagnosis of peribronchial lesions\textsuperscript{8} and is also an approach which enables the identification of PE as bronchi are always accompanied by a pulmonary artery.\textsuperscript{9-12} Therefore, we considered that the EBUS-guided transbronchial needle aspiration (TBNA) needle could be an effective means of reaching PE transbronchially and devised a novel therapeutic approach for PE using EBUS-TBNI.\textsuperscript{13, 14} EBUS-TBNI can be a novel therapeutic approach for more accurate drug delivery by selecting the injection site within the thrombus under real-time ultrasound guidance in order to dissolve clot effectively.

The safety of performing EBUS-TBNA through the pulmonary artery has previously been reported.\textsuperscript{15-19} Regarding pulmonary artery punctures using EBUS-TBNA needle based on skilled technique and adequate anatomical knowledge, the possibility of bleeding is considered to be low. Accidental puncture of the pulmonary artery by a trainee can occurs in practice, but major bleeding is extremely uncommon as evidenced by reports that bleeding is a rare complication associated
with EBUS-TBNA. Therefore, the approach of puncturing the pulmonary artery itself may be considered as an acceptable technique, but it is unclear whether t-PA injection using EBUS-TBNI is safe. To clarify this point, we have reported on t-PA injection using EBUS-TBNI in the porcine PE model. No complications were observed for one week after t-PA injection using EBUS-TBNI. This was evaluated by bronchoscopy and examination of the resected lungs, not by CT scans.

In this study, using contrast-enhanced CT scans, hemodynamic parameters, and blood test (including coagulation over time), we evaluated the effectiveness in clot dissolution by t-PA injection using EBUS-TBNI and assessed the safety of this technique.

Methods:

All experiments were approved by the University Health Network Animal Care Committee (Animal Use Protocol 5928.5.2) on August 10th, 2021. Six Yorkshire pigs (mean weight 31.9 ± 2.9 kg; Caughell Farms Ltd) were used as the porcine PE model. EBUS and EBUS-TBNI were performed with a convex probe EBUS bronchoscope (BF-UC180F; Olympus Corporation) with a dedicated ultrasound processor (EU-ME1; Olympus Corporation). A 21 or 22 gauge ViziShot 2 EBUS-TBNA needle (NA-U401SX-4021-A/NA-U401SX-4022-A; Olympus Corporation) was used for clot injection and a 25 gauge ViziShot 2 EBUS-TBNA needle (NA-U401SX-4025N; Olympus Corporation) was used for t-PA injection.

Animal Model

Methods of anesthesia and monitoring during the procedures were used as previously described. Under general anesthesia, the pig underwent intubation with an 8.5 mm endotracheal tube and then was set on a pressure-controlled ventilator. An intravenous injection
catheter was placed at the pig ear and a Swan-Ganz catheter (Synthetic ControlCath TD catheter C146F7; Edwards Lifesciences Corporation) was placed into the jugular vein via percutaneous sheath introducer kit (Arrow Percutaneous Sheath Introducer Kits 9 Fr AK-09903-CDC; Teleflex Incorporated). Maintenance of anesthesia took place with monitoring of blood pressure, oxygen saturation, end tidal CO₂, ECG, and rectal temperature using a multiparameter monitor (BM3Vet touch; Bionet Co.). Pulmonary artery pressure (PAP) was measured using a pressure transducer (TruWave Disposable Pressure Transducer Standard Kits PX284; Edwards Lifesciences).

The experimental schedule on day 1 is described in figure 1. Sixty mL of autologous blood was withdrawn and maintained at room temperature for 2 hours in order to create stable clots contained in syringes. After 2 hours, the serum was discarded and a total of 30 mL of the clots was dispensed to 1 mL syringes for clot injection. Fifteen mL of clot was injected into bilateral pulmonary arteries. The first 15 mL of clot was injected into the right pulmonary artery and the second 15 mL of clot was injected into left pulmonary artery, through the EBUS-TBNA needle (Video 1). Since our previous results showed that the amount of thrombus formation in the pulmonary artery varied among individuals, we decided to conduct the experiment with an aligned volume of clots in order to create PE. Clot injection started from the most distal main pulmonary artery, limited only by the reach of EBUS scope, since the clot is easily dispersed throughout the segmental pulmonary arteries when clot injection is performed in the main pulmonary artery starting from the central side. Once the injected clot reached the tip of EBUS-TBNA needle on EBUS, the puncture site was moved proximal to the pulmonary artery, and the clot was injected at three points in total to create a continuous thrombus. Thrombolytic agent, t-PA was administered 2 hours post clot injection.

We designed two types of thrombolytics treatment method for the porcine PE model. The
first treatment is an injection of 25 mg of t-PA, dissolved in 25 mL distilled water, into a PE using EBUS-TBNI (TBNI group). Fifty mg of t-PA was used for bilateral PEs in a pig in total. As we reported previously, t-PA was then injected directly into the clots using EBUS-TBNI by a 25 gauge needle. Clots were punctured in 3 places sequentially from distal to proximal, with one-third of 25 mg t-PA administered at each puncture site (Video 2). The second treatment is a systemic administration of 100 mg t-PA, delivered intravenously over 2 hours (Systemic group as a control) conforming to the suggested treatment in the ACCP and the ESC guidelines.\textsuperscript{1-3} Hemodynamic parameters, blood collection, and contrast enhanced CT scans underwent at the time of (1) baseline as control, (2) pre t-PA, (3a) 30 minutes post t-PA in TBNI group, and (3b) 2 hours post t-PA in Systemic group on day 1. Only the hemodynamic parameters at 30 minutes post t-PA systemic administration in Systemic group were additionally collected. Those data were collected on (4) day 3 and (5) day 7. On day 7, the pigs were euthanized 5 minutes after 10,000 U (10 mL) heparin administration and underwent bilateral pneumonectomy to measure the residual clots in both of pulmonary arteries.

Measurement of Clot Volume

Regarding the analysis of the residual clot volume, we collected the clot volumes separately from each pulmonary artery in one pig, so we used 3 pigs for each group, meaning 6 residual clot volumes were evaluated from each group. The clot volumes in pulmonary arteries were measured using the 3D slicer software (version 5.0.3 r30893/7ea0f43).\textsuperscript{24} This is an open-source software application for medical image computing. The 3D Slicer application framework provides standard Digital Imaging and Communications in Medicine (DICOM) import and export, 2D image slicing and 3D visualization and quantification. In this study, Segment Editor and Quantification module
were conducted to measure the clot volumes. The clots were traced on each slice of CT images. The 3D slicer software automatically generates a 3D model using these traced areas and calculates the amount of clot volumes. Regarding the residual clots on day 7 confirmed by pneumonectomy, the volume was calculated using the truncated cone formula we previously reported\textsuperscript{23} because we were not able to clearly determine the clot volume based on the CT due to the lack of sensitivity in this modality.

\section*{Statistical Analysis}

Summary statistics are expressed herein as mean ± standard error of the mean. A two-factor repeated measures ANOVA was conducted to compare the hemodynamic parameters and the blood test between the groups over time. \(P\)-values are indicated for pairwise comparisons of interest and were computed by using Sidak’s multiple comparison test between treatment groups and by using Tukey’s multiple comparisons test between time points when a significant difference was shown in the result of the ANOVA. The residual clot volume and percent change of clot volume were also conducted by a two-factor repeated measures ANOVA. Clot volume decrease per hour was compared between treatment groups using the Mann-Whitney U test. The Wilcoxon signed-rank test was used for comparison between each time point in one treatment group. A \(p\)-value of less than 0.05 was considered statistically significant (GraphPad Prism ver. 9.5.0; GraphPad Software).

\section*{Results}

Bilateral clot injections were performed on 6 pigs: 3 for Systemic group; 3 for TBNI group. In all cases, thrombus formation was successful, and we were able to measure the hemodynamics
parameters, blood tests, and acquire the enhanced-contrast CT scans. All pigs survived until euthanasia on day 7. The results of the hemodynamic parameters and the blood tests for both groups are summarized in Figure 2. In both groups, PAP increased immediately after clot injection and decreased gradually over 2 hours. The results of sPAP and dPAP showed significant differences in the time factor by two-factor repeated measures ANOVA, respectively (sPAP, F = 13.67, p < 0.0001; dPAP F = 11.9, p < 0.0001). However, the result between Systemic group and TBNI group did not show significant differences (Figure 3A, B). For pairwise comparisons of sPAP between each time point in TBNI group, there were some significant differences between the time points; in Systemic group, there were also some significant differences between the points (Supplemental figure 1A, B). The pairwise comparisons of dPAP between each time point showed similar results as well (Supplemental figure 1C, D).

The average of the d-dimer plateaued at a maximum at the post-t-PA timepoint and then decreased and returned to baseline in both groups on day 3. The average of d-dimer showed significant differences in the time factor by two-factor repeated measures ANOVA (F = 4.84, p < 0.01). There was no significant difference between the groups (Figure 3C). The average of the fibrinogen also didn’t show any significant differences in the time factor and in the group factor (Figure 3D). The other hemodynamic parameters and blood tests were also measured. Only the average of pH showed significant differences in the group factor by two-factor repeated measures ANOVA (F = 11.05, p < 0.05) (Figure 4).

The evaluations of residual clot volumes are shown in figure 5A (mm³: Systemic group 611.0 ± 234.4 vs TBNI group 880.1 ± 799.2 at pre t-PA; Systemic group 221.8 ± 139.6 vs TBNI group 50.3 ± 28.4 at day 3). For the residual clot volume, a two-way repeated measures ANOVA was statistically significant for differences in the time factor (F = 11.3, p < 0.01). For pairwise
comparisons of the residual clot volume between each time point in TBNI group, there were significant differences between pre t-PA/day 3 ($p < 0.05$). There was no significant difference between both groups. Looking at the percent change of clot volumes, normalizing each residual clot volume at the time point of post clot as 100%, the average of % change of the residual clot volumes also showed significantly differences in the time factor ($F = 214.2, p < 0.0001$) and in the group factor ($F = 6.609, p < 0.05$) by a two-factor repeated measures ANOVA (Figure 5B). For pairwise comparisons of the percent change of clot volumes between Systemic group and TBNI group, there was a significant difference ($%$: Systemic group 36.6 ± 22.6 vs TBNI group 9.6 ± 6.1, $p < 0.01$). Measuring the elapsed time from the time record of the contrast enhanced CT scan, this measured time was used to calculate the clot volume decrease volume per hour. Considering the elapsed time, the average decrease of clot volume per hour at pre t-PA to post t-PA was higher in TBNI group than in Systemic group (mm$^3$/hour: Systemic group 68.1 ± 68.1 vs TBNI group 256.8 ± 148.1, $p < 0.05$) (Figure 5C). The speed of clot dissolution was about 3.8 times faster in TBNI group than in Systemic group at t-PA to post t-PA. Comparing two terms in each group separately, only the TBNI group showed a significant decrease between two terms (mm$^3$/hour: in TBNI group, from pre t-PA to post t-PA 256.8 ± 162.2 vs from post t-PA to day 3 11.3 ± 15.0, $p < 0.05$). On day 7, there was one residual clot in TBNI group, and 3 clots in Systemic group, respectively. There was no significant difference in the average of residual clot volumes on day 7 between Systemic group and TBNI group (mm$^3$: Systemic group 22.4 ± 29.9 vs TBNI group 1.6 ± 3.9, $p = 0.18$) (Supplemental figure 2).

Regarding complications, one pig in TBNI group developed stridor on postoperatively day 1. We believe this is due to failed intubation. Therefore, it was determined to be laryngeal edema caused by the damage of failed intubation. The stridor improved after administrating
Dexamethasone (0.1 mg/kg) intramuscularly, but it lasted until day 7. No hemorrhage caused by t-PA administration and EBUS-TBNI was observed intracranially, intrathoracically, or intraperitoneally on any contrast-enhanced CT images in either group. Bronchoscopy views did not show any bleeding during procedures (Video 3). In the systemic group, one pig had a large consolidation in right lower lobe, which spread between right main bronchus and dorsal lower lobe. These radiological findings revealed atelectasis by pathological assessment.

Discussion

This study showed the effectiveness in clot dissolution and the safety of t-PA injection using EBUS-TBNI for the porcine PE model (Figure 6).

Regarding the dissolving of the clot via t-PA injection using EBUS-TBNI; this treatment was effective immediately after t-PA administration, correlating with results of previous studies. In particular, as shown in Figure 5C, the clot volume decrease per hour clearly shows the immediate effect after t-PA administration in TBNI group, with a dissolution rate of 256.8 mm³/hr (compared to 68.1 mm³/hr in Systemic group). This result suggests that this treatment method can be applied to massive PE in emergency situations. However, t-PA injection using EBUS-TBNI was not sufficiently effective to reduce the average PAP at 30 minutes post t-PA (Figure 3A, B).

There were no significant differences in PAP between both groups, however focusing on day 3, there were differences between both groups. This result becomes interesting when considering clinical trials using this treatment. In our model, the porcine thrombus filled the pulmonary artery in the caudal lung lobe sequentially from the periphery, which suggests that the clot must be dissolved all the way down to the periphery to release the increased PAP. This may explain why the results of PAP were not as effective as the rate of thrombus dissolution. In the case of normal
acute PE at high risk, the thrombus is often stuck in the central side of pulmonary artery, and it is important to dissolve the thrombus in the central pulmonary artery or dislodge it quickly into the peripheral pulmonary artery, where it would pose a smaller risk. In this respect, the superiority of t-PA injection using EBUS-TBNI is evident in the thrombus volume data, so we do not believe that the fact there were no significant differences in PAP is a problem.

It has been reported that a decrease of fibrinogen is associated with bleeding during t-PA treatment. No significant differences between the two groups were observed, nor were there any significant differences from baseline. It could be considered a reasonable result as no bleeding complications occurred. The values of d-dimer also showed no significant differences between two groups, however the d-dimer increased immediately after clot injections so that this result was also reasonable as d-dimer is used to exclude the diagnosis of PE. Other than PAP, the hemodynamic monitoring was not significantly different between the two groups. This might be due to the pigs being under general anesthesia and ventilator management with tracheal intubation, and therefore, the systemic management was adjusted as needed.

We previously reported that a minimum of 10 mg t-PA injection using EBUS-TBNI can provide maximum efficacy on PE as well as 25 mg t-PA injection using EBUS-TBNI. Administration of t-PA has dose-dependent bleeding risk in general. Since one of the purposes in this study was to confirm the safety, experiments were conducted using a higher dose of 25 mg t-PA injection EBUS-TBNI. Three pigs were used for each group in this study, therefore the conclusion on the safety of t-PA injection using EBUS-TBNI is limited by the small sample size.

In our previous experiments, no intra-airway bleeding was observed in 6 pigs during a one-week survival model and in the 8 pigs with non-survival model. Combining the previous results with the current experiment, a total of 75 punctures of t-PA injection using EBUS-TBNI have been
performed on 26 pulmonary arteries in 17 pigs. We have never observed any active bleeding from pulmonary artery by bronchoscopy (Table 1) although there were no CT scans and hemodynamic parameters collected at that time. Taking this into account, we presume that major bleeding in the airway is unlikely to occur in t-PA injection using EBUS-TBNI.

The first key advantage of t-PA injection using EBUS-TBNI for PE is that EBUS can evaluate the location of PE in real time, and unlike the t-PA systemic administration or USAT, t-PA using EBUS-TBNI can be administered at a pinpoint within the thrombus. t-PA is only effective at dissolving clots when it is present with the clots at the same time. Therefore, by injecting t-PA directly into the thrombus, it might be possible to achieve its effect efficiently and to minimize the total dosage of t-PA administered. Although direct placement of a catheter in the pulmonary artery to inject t-PA should be considered to increase the effect of t-PA, guidelines point out that there may be higher complications than systemic administration. Therefore, that technique was not considered in this study. The second key advantage of t-PA injection using EBUS-TBNI is its ease of adoption into current practice. EBUS-TBNA and USAT both require training for proficiency before implementation, but unlike USAT, EBUS system are usually already equipped in core hospitals that can treat acute PE.

This study has several important limitations. First, the number of animals used in this study was small. However, looking at the difference in treatment efficacy between t-PA systemic administration and t-PA injection using EBUS-TBNI, there is a significant difference in the speed of clot dissolution, which suggests that even with only three cases, sufficient data was obtained as described above. On the other hand, to the point of safety, no conclusions could be drawn from this study alone without considering the previous studies. Since this was an animal study, it is difficult to guarantee the safety, so the decision should take past studies into consideration. Second,
since this experiment was conducted in a porcine model, it is important to note the difference in anticoagulation between human and pig. Although reported in vitro, porcine blood has similar maximum coagulation firmness\textsuperscript{27}, and has prolonged clot lysis by t-PA compared to humans.\textsuperscript{28} However, as our previous\textsuperscript{13} and current studies have shown, the response of t-PA to PE is still effective in vivo. This suggests the possibility of further reduction of the t-PA dose when t-PA injection using EBUS-TBNI is adapted to humans. Regarding intubation complication; although intubation was performed for the pig experiment, complications due to intubation are not expected to cause any issues in a clinical setting as this procedure would be carried out under conscious sedation without intubation. Third, this study did not evaluate the safety of using parenteral anticoagulation concurrently or after t-PA injection using EBUS-TBNI. It is recommended that the acute PE at high risk receives a parenteral anticoagulation during or after t-PA administration.\textsuperscript{1}\textsuperscript{3} Since it has been noted that t-PA administration during systemic heparinization is likely to increase the risk of bleeding, careful consideration should be given regarding the timing of heparin initiation when this technique is actually used for treatment. However, this technique is apparently less invasive than the pulmonary embolectomy. Therefore, we assume this technique followed by the systemic heparinization could be acceptable in case heparinization starts 4-6 hours later because theoretically t-PA is rapidly metabolized. Fourth, this experiment is only effective in quickly dissolving thrombus, but it is not clear whether it truly improves cardiac burden such as RV dysfunction and prognosis. However, since it is difficult to evaluate this in animal studies, it will become clear in future clinical trials.

\textbf{Conclusions}

This study has revealed that t-PA EBUS-TBNI can be an effective procedure to safely
dissolve acute massive PE. Its dissolving effect is more immediate than systemic administration of t-PA. This efficacy data suggests a potential role for a clinical trial.
References


25. Kaufman C, Kinney T, Quencer K. Practice Trends of Fibrinogen Monitoring in


Table 1. The total number of pigs used, pulmonary arteries used, and punctures done to evaluate t-PA injection using EBUS-TBNI over 3 studies.

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Figure legends

Figure 1. This illustration shows the procedures on day 1. The green frame shows the collection time points for the hemodynamic parameter, the blood test, the CT scans, and the EBUS images. The red frame shows the time points for clot injection or t-PA administration. Note that the Systemic group hemodynamic parameters were collected 30 minutes and 2 hours after t-PA administration. The other data: the blood tests, the EBUS images, and the CT scans were collected 2 hours post t-PA administration. Systemic group: t-PA systemic administration; TBNI group: t-PA injection using EBUS-TBNI.

Figure 2. The changes in each hemodynamic parameter at each time point in Systemic group and TBNI group. The average changes of (A) systolic BP, (B) diastolic BP, (C) heart rate, (D) SpO₂, and (E) end-tidal CO₂ were shown at each time point for each group.

Figure 3. The changes in pulmonary artery pressure (PAP), d-dimer, and fibrinogen at each time point in Systemic group and TBNI group. The average changes of (A) systolic PAP, (B) diastolic PAP, (C) d-dimers, and (D) fibrinogen are shown at each time point for each group.

Figure 4. The changes in each blood test at each time point in Systemic group and TBNI group. The average changes of (A) pH, (B) HCO₃, (C) pO₂, (D) pCO₂, (E) PT, and (F) PTT were shown at each time point for each group.
Figure 5. Analysis of the residual clot volumes. (A) The comparison of the residual clot volumes between Systemic group and TBNI group at pre t-PA and on day 3. (B) The percentage change of clot volumes on day 3 compared to pre t-PA. (C) Amount of clot volume decrease per an hour from pre t-PA to post t-PA and from post t-PA to day 3. *, p < 0.05; **, p < 0.01. The lower and upper borders of the box represent the lower and upper quartiles. The middle horizontal line represents the median. The lower and upper whiskers represent the minimum and maximum values of non-outliers. Each value is plotted in the same color.

Figure 6. (Graphical abstract)

Summary of this study on the safety and effectiveness of endobronchial ultrasound-guided transbronchial needle injection (EBUS-TBNI) in a porcine survival model. A pig model of pulmonary embolism (PE) was created by injecting clots into the pulmonary artery of healthy pigs under EBUS guidance. One group received 100 mg t-PA systemic administration to treat PE (Systemic group); the other group received each 25 mg t-PA injection into bilateral PEs using EBUS-TBNI (total of 50 mg t-PA). Hemodynamic parameters, blood tests, and CT scans were collected to evaluate the clot dissolving. In TBNI group, the speed of dissolving clot was significantly higher than in Systemic group. There were no complications in both groups. Twenty five mg t-PA injection using EBUS-TBNI can be a safe and effective procedure to treat acute massive PE. The promising efficacy data suggest a potential role for clinical practice.
Videos

Video 1. Clot injection into the pulmonary artery by EBUS-TBNA needle.

Video 2. Administration of t-PA using EBUS-TBNI.

Video 3. Bronchial view 30 minutes after t-PA injection using EBUS-TBNI.
Supplemental figures

Supplemental figure 1. The changes in systolic PAP and diastolic PAP in Systemic group and TBNI group. The average changes of (A) systolic PAP in Systemic group, (B) systolic PAP in TBNI group, (C) diastolic PAP in Systemic group, (D) diastolic PAP in TBNI group. *, $p < 0.05$; **, $p < 0.01$; ***, $p < 0.001$.

Supplemental figure 2. The comparison of the residual clot volumes between Systemic group and TBNI group on day 7. ns, not significant.
Assessment effectiveness and safety of Thrombolytic Therapy to Pulmonary Emboli (PE) by Endobronchial Ultrasound-guided Transbronchial Needle Injection (EBUS-TBNI) in a Porcine Survival Model

Pig PE model

100mg t-PA systemic administration in 3 pigs

Measurements of hemodynamic parameters, blood tests, and dissolved clot volumes until day 7

25mg t-PA injection using EBUS-TBNI in 3 pigs

Representative contrast-enhanced CT images

This study has revealed that t-PA EBUS-TBNI can be an effective procedure to safely dissolve acute massive PE.
Assessment of Safety and Feasibility of Thrombolytic Therapy to Pulmonary Emboli by Endobronchial Ultrasound-guided Transbronchial Needle Injection in a Porcine Survival Model

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